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## NEW CHLORINE-CONTAINING PHENOLOID FROM *CURCULIGO CAPITULATA*

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A new chlorine-containing phenoloid, named capitulatin A, has been isolated from the rhizomes of *Curculigo capitulata*. Its structure was established as 2,4-dichloro-3-methyl-5-hydroxy-6-methoxyphenol- $\beta$ -D-xylopyranosyl (1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (**1**) on the basis of the spectral data and chemical evidence.

**Keywords:** *Curculigo capitulata*; Hypoxidaceae; Chlorine-containing phenoloid; Capitulatin A

### INTRODUCTION

The herb *Curculigo capitulata* (Lour.) Ktze, used as a tonic and a medicine for treating dysmenorrhea and rheumatism [1], is widely distributed in Southern and Southwestern China, Taiwan (China), Malaysia, India and Australia. Various compounds, including phenanthropyran [1], norneolignan and phenols [2], have been isolated from this plant. In this paper a new chlorine-containing phenoloid (**1**) was isolated from the rhizomes of *C. capitulata*. Chlorine-containing phenoloids of this kind, reported previously from the same genus, *C. orchioides* [3–5], are very scarce. Here we report the structural elucidation of **1**.

### RESULTS AND DISCUSSION

Compound **1** was obtained as colorless needles (MeOH). Its HRFAB<sup>−</sup>MS gave a quasi-molecular ion at  $m/z$  515.0784  $[M - 1]^{-}$  and fragment ions at  $m/z$  517.0748  $[M - 1 + 2]^{-}$ , 519.0732  $[M - 1 + 4]^{-}$ , in which the relative abundance ratio for  $[M - 1]:[M - 1 + 2]:[M - 1 + 4]$  was 9:6:1, indicating that **1** contains two chlorines

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and a molecular formula of  $C_{19}H_{26}O_{12}Cl_2$ , which was confirmed by the  $^{13}C$  NMR data. In the  $^{13}C$  NMR spectrum, carbon signals at  $\delta$  145.1s, 144.5s, 142.5s, 125.9s, 125.6s and 125.0s confirmed the presence of hexa-substituted aromatic ring with three  $-OR$ ,  $-C$  and two  $Cl$  groups. Furthermore, chemical shifts at  $\delta$  17.8 and 61.1 in  $^{13}C$  NMR spectrum are in accordance with  $\delta$  2.69 (s, 3H) and 4.13 (s, 3H) in the  $^1H$  NMR spectrum and indicate that **1** contains one  $CH_3$  and one  $OCH_3$  group. The correlation peaks between C-3 ( $\delta$  125.9) and protons of  $CH_3$ , C-6 ( $\delta$  145.1) and protons of  $OCH_3$  in the HMBC spectra of **1** (Fig. 1) confirm that  $CH_3$  is linked with C-3 and  $OCH_3$  with C-6. Two quaternary carbon signals, at  $\delta$  125.6 and 125.0, correlate with the protons of  $CH_3$  in HMBC, indicating the attachment of two chlorines in C-2 and C-4. In the  $^{13}C$  NMR spectrum the carbon signals at  $\delta$  105.2, 74.8, 77.6, 71.1, 66.9 and  $\delta$  106.5, 75.3, 77.5, 70.9, 77.9, 69.4 indicated the presence of one xylose and 6-substituted glucose. Acid hydrolysis of compound **1** showed that it contains glucose and xylose. In the FAB-MS of **1**, the ion peaks at  $m/z$  383 $[M - 1 - xyl]^-$  and 221 $[M - 1 - xyl - glc]^-$  suggested the presence of the two sugar moieties. Two doublets at  $\delta$  5.10 ( $J = 7.60$  Hz) and 4.57 ( $J = 7.30$  Hz) in the  $^1H$  NMR spectrum also indicate the presence of a glucose unit and a xylose unit in **1**. From the coupling constants of the anomeric protons and the  $^{13}C$  NMR chemical shifts, the two sugar moieties must be  $\beta$ -type sugars. The other positions of the two sugars were confirmed by  $^1H$ - $^1H$  COSY, HMQC-TOCSY and NOE spectroscopy. The cross signals at  $\delta$  142.5 (C-1)/2.69 (proton of  $CH_3$ ),  $\delta$  142.5 (C-1)/5.10(H-1') and  $\delta$  69.4 (C-6')/4.57(H-1'') in the HMBC spectrum reveal a 1-*O*-glucoside and 1 $\rightarrow$ 6 linkage of the two sugars moieties. In the NOESY spectrum of **1**, cross-peaks were observed between the methoxyl protons (4.13) and H-6'(4.30) of the glucose unit, indicating that the two quaternary carbons, attached by methoxyl group and glucose unit, are adjacent. On acetylation of **1**, the saponin acetate was subjected to FAB-MS analysis, and showed  $m/z$  515 + 7  $\times$  42 $([M - 1]^- + 7Ac - 7H)(809)$ , which indicates the presence of an hydroxy group linked with the aromatic ring. A downfield chemical shift at  $\delta$  144.5 was due to C-5 linked with the hydroxy. From the above-mentioned evidence, compound **1** was determined to be 2,4-dichloro-3-methyl-5-hydroxy-6-methoxyphenol- $\beta$ -D-xylopyranosyl (1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (Fig. 1).

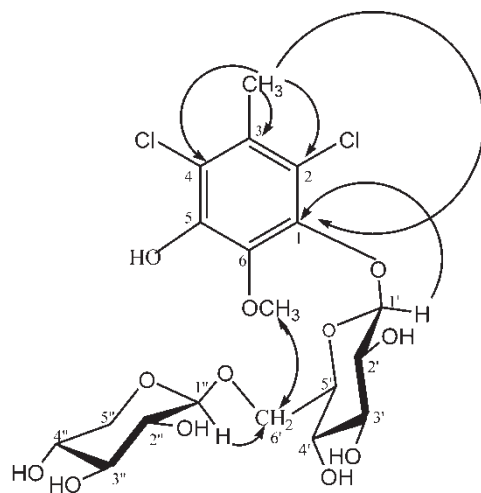


FIGURE 1 Structure of compound **1** ( $\rightarrow$  HMBC correlations;  $\leftrightarrow$   $^1H$ - $^1H$  NOESY correlations).

## EXPERIMENTAL

### General Experimental Procedures

The mp was determined on an XRC-1 micro melting point apparatus and is uncorrected.  $[\alpha]_D$  was determined with a JASCO-20. IR spectra were recorded on a Bio-Rad FTS-135 spectrometer with KBr pellets. UV spectra were recorded on a UV 210A spectrometer. 1D and 2D NMR spectra were run on Bruker DRX-500 instruments with TMS as internal standard using CD<sub>3</sub>OD as solvent. The FAB-MS was carried out on a VG Auto Spec-3000 spectrometer. TLC was carried on silica gel G (MEIJING) precoated plates. Spots were detected by spraying with 5% sulfuric acid–ethanol solution followed by heating.

### Plant Material

Rhizomes of *C. capitulata* were collected from the west garden of Xi Shuang Ban Na Botanical Garden and identified by Professor Zhou Jun of the Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen was deposited in the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences.

### Extraction and Isolation

The air-dried and powdered rhizomes of *C. capitulata* (3 kg) were extracted with 85% EtOH (3 × 20 L) at room temperature, and the combined extracts were evaporated *in vacuo*. The residue was suspended in H<sub>2</sub>O and then passed through a D101 resin column to eliminate sugars; the column was then eluted with 95% EtOH. The EtOH eluent was concentrated *in vacuo* to give a residue (240 g) that was chromatographed on silica gel column (200–300 mesh) with CHCl<sub>3</sub>–MeOH (7:2) to give 8 fractions. Fraction 4 was subjected to column chromatography over silica gel eluted with CHCl<sub>3</sub>–MeOH (10:1.5) to afford **1** (12 mg, yield 0.05%).

### 2,4-Dichloro-3-methyl-5-hydroxy-6-methoxyphenol-β-D-xylopyranosyl(1-6)-β-D-glucopyranoside (**1**)

Colorless needles (MeOH), mp 174–178°C,  $[\alpha]_D^{21} - 36.67$  (*c* 0.30, MeOH); IR (KBr)  $\nu_{\max}$  (cm<sup>-1</sup>): 3419, 2927, 1634, 1467, 1072, 1045, 994; UV(MeOH)  $\lambda_{\max}$  (nm): 286, 205. <sup>1</sup>H NMR (CD<sub>3</sub>OD, MHz)  $\delta$ : 2.69 (3H, s, CH<sub>3</sub>-3), 4.13 (3H, s, CH<sub>3</sub>O-6), 4.57 (1H, d, *J* = 7.30 Hz, H-1''), 5.10 (1H, d, *J* = 7.60 Hz, H-1'); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 125 MHz) data are shown in Table I. Molecular formula C<sub>19</sub>H<sub>26</sub>O<sub>12</sub>Cl<sub>2</sub> (negative HRFAB MS 515.0732; calcd. 515.0723).

### Acid Hydrolysis of **1**

Compound **1** was dotted on a silica gel G plate, placed and hung in a sealed glass vessel with concentrated HCl (ca. 1 ml) at 70°C for 1 h for hydrolysis and then cooled for a few minutes; the plate was taken out, and the HCl volatilized with a ventilator. Authentic sugars were dotted to the plate, which was then developed with n-butanol–pyridine–water (6:4:3), and 5% sulfuric acid–ethanol solution used as spray reagent, followed by heating at 120°C. From compound **1** glucose and xylose were detected; *R<sub>f</sub>*: glucose 0.40, xylose 0.52.

TABLE I  $^{13}\text{C}$  NMR spectral data of compound **1** ( $\text{CD}_3\text{OD}$ , 125 MHz)

	Aglycone		Sugars
1	142.5 s	Glucose	
2	125.0* s	1'	106.5 d
3	125.9 s	2'	75.3 d
4	125.6* s	3'	77.5 d
5	144.5 s	4'	70.9 d
6	145.1 s	5'	77.9 d
—CH <sub>3</sub>	17.8 q	6'	69.4 t
—OCH <sub>3</sub>	61.1 q	Xylose	
		1''	105.2 d
		2''	74.8 d
		3''	77.6 d
		4''	71.1 d
		5''	66.9 t

\* Data are exchangeable.

### Acetylation of **1**

Compound **1** (1 mg) was dissolved in  $\text{Ac}_2\text{O}$ –pyridine (1:0.5) in a sealed micro-tube. After reacting at 60–70°C for 6 h, the acetate of **1** was subjected to FAB-MS analysis.

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